

ACHILLICIN, THE FIRST PROAZULENE FROM *ACHILLEA* *MILLEFOLIUM*

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Abstract—Achillicin, the major proazulene (prochamazulene) of *Achillea millefolium* L. ssp. *collina* Becker has been isolated and identified as 8-acetoxyartabsin. It is the first proazulene reported for the genus *Achillea* and has not been found previously in plants.

INTRODUCTION

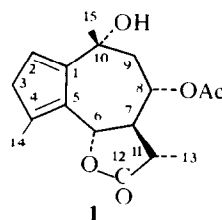
For many years, *Achillea millefolium* (yarrow, milfoil) has been known to yield proazulene-rich essential oils. Despite extensive investigations into its constituents [1–11], identification of the natural proazulenes has not been reported. Matricin, a proazulene from chamomile (*Matricaria chamomila*) has been reviewed [12] as occurring also in some *Achillea* species (*A. collina* Becker, *A. crithmifolia* W. et K., *A. pannonica* Scheel, *A. stricta* Schleid. In the paper cited [13] however, the occurrence of proazulenes in general, and not that of matricin had been stated. Similarly, the suggestion [14] that chamazulene carboxylic acid is a natural proazulene of the genus *Achillea* proved erroneous [15].

Now we report the isolation and structure elucidation of achillicin, the first natural proazulene identified for the genus *Achillea*. This compound has not been previously found in plants.

RESULTS AND DISCUSSION

Accurate mass measurements gave the molecular formula of achillicin as $C_{17}H_{22}O_5$ (306.145, requires 306.146) and MS indicated the presence of an acetoxy function (m/e 246.124, $C_{15}H_{18}O_3^+$, requires 246.126, M-60) and a OH group (m/e 228.113, $C_{15}H_{16}O_2^+$, requires 228.115, M-60-18). These were also confirmed by IR (1727 cm^{-1} , CO and 3453 cm^{-1} , OH) which, in addition, disclosed the presence of a saturated γ -lactone (1773 cm^{-1}), a tertiary OH group (1153 cm^{-1}) and two conjugated double bonds (1613 cm^{-1}). Treatment of achillicin in boiling 5% NaOH yielded chamazulene carboxylic acid (identified by IR, MS, PMR and ^{13}C NMR spectra and by transformation into chamazulene) which established the guaiazulene skeleton.

Conclusive information on the structure and stereochemistry of achillicin as **1** was inferred from PMR and ^{13}C NMR data (Tables 1 and 2). Disregarding the acetoxy group, the PMR data are in excellent agreement with values reported for artabsin [16–18]. 1H - $\{^1H\}$ double resonance experiments defined the positions of the conjugated double bonds and the acetoxy group. The stereochemistry at C-6, C-7, C-8 and C-11 was readily elucidated by 1H - $\{^1H\}$ coupling constants (see Table 1).



The relative configuration at C-10 was inferred from consideration of ^{13}C chemical shifts indicating an equatorial orientation for the C-10 Me group (lack of 1,3 diaxial steric interactions).

EXPERIMENTAL

IR: KBr; MS: 70 eV, probe, source temp. 130°; PMR and ^{13}C NMR: 100.1 and 25.16 MHz, $CDCl_3$.

Isolation of **1**. The air-dried (40 g) flowerheads of *A. mille-*

Table 1. PMR data (δ ppm from TMS) for achillicin

2-H	6.09	C-4Me	2.19
3-H	2.90	C-10Me	1.62
6-H	5.50	C-11Me	1.30
8-H	5.52	—OH	1.73
9-H _{ax}	1.66	O—CO—Me	2.09
9-H _{eq}	2.28		
11-H	2.62		

J(Hz): 2,3 = 1.5; 3,6 = 1.5; 6,7 = 10; 6,C-4Me = 2; 7,8 = 11; 8,9_{ax} = 10.5; 7,11 = 11; 8,9_{eq} = 4; 9_{ax},9_{eq} = 14; 11,C-11Me = 7.

Table 2. ^{13}C NMR chemical shifts (δ ppm from TMS) for achillicin

C-1	137.85	C-9	46.50
C-2	124.54	C-10	69.40
C-3	45.58	C-11	55.94
C-4	149.84	C-12	178.20
C-5	134.04	C-4Me	14.90
C-6	78.24	C-10Me	30.38
C-7	40.50	C-11Me	15.50
C-8	73.46	O—CO—Me	21.16
		O—CO	169.96

folium L., ssp. *collina* Becker collected in summer 1976 near Budapest, Hungary, were extracted with Et₂O (1.5 l) and made alkaline with few drops of NH₄OH. Hexane (40 ml) and H₂O (400 ml) were added to the extract which was then evapd in a rotary evaporator. After filtration, the aq. filtrate was extracted with CHCl₃ (3 × 100 ml). The solvent was evapd under red. pres. and the resulting extracts were separated by column chromatography (Si gel 40, C₆H₆-EtOAc, hexane-EtOAc-CHCl₃). The fractions were analysed by TLC (Si gel G) using hexane-EtOAc-CHCl₃ (9:5:6) as solvent. Components were visualized by spraying with EP reagent [20], followed by heating for 10 min at 100° and subsequent spraying with EtOH for colour enhancement [15]. On removal of the solvent, the most EP-positive chromatographic fractions afforded crystalline achillicin (*R_f* = 0.3).

Conversion of 1 to chamazulene carboxylic acid. A few mg of 1 were boiled in 5% aq. NaOH (5 ml) for 15 min, cooled, neutralized with 2N H₂SO₄ and extracted with hexane. Evapn gave pure chamazulene carboxylic acid.

REFERENCES

1. Falk, A. J. (1973) *Diss. Abstr. Int. B* **34**, 2537.
2. Kaneko, H., Naturo, Sh. and Takahashi, Sh. (1971) *Phytochemistry* **10**, 3305.
3. Kasymov, Sh. Z. and Sidyakin, G. P. (1972) *Khim. Prir. Soedin.* 246.
4. Linde, H. H. A. and Ragal, M. S. (1967) *Helv. Chim. Acta* **50**, 1961.
5. Neshta, I. D., Rybalko, K. S., Konovalova, O. A. and Ivanenko, O. F. (1976) *Khim. Prir. Soedin.* 395.
6. Neshta, I. D. and Kaloshina, N. A. (1972) *Khim. Prir. Soedin.* 652.
7. Romo de Vivar, A. and Olmo, F. (1968) *Rev. Soc. Chim. Mex.* **12**, 212.
8. Smolenski, St. J., Bell, C. L. and Bauer, L. (1967) *Lloydia* **30**, 144.
9. Tillya'ev, P., Khamatov, Kh. Kh., Prikhamedov, I. and Talipova, M. A. (1973) *Rastit. Resur.* **9**, 58.
10. White, E. H. and Winter, R. E. K. (1963) *Tetrahedron Letters* 137.
11. Winter, R. E. K. and White, E. H. (1972) *Quimica* **68**, 827.
12. Yoshioka, H., Mabry, T. J. and Timmermann, B. N. (1973) in *Sesquiterpene Lactones*. University of Tokyo Press, Tokyo.
13. Kotilla, E. (1959) *Orvosi Szemle* **5**, 308.
14. Tyihák, E. and Vágúfalvi, D. (1967) *Planta Med.* **15**, 269.
15. Cuong, B.N., Verzar-Petri, G. and Tamas, J. (1977) *Proc. 17th Hung. Annu. Meet., Biochem. Kecskemét* 127.
16. Geissman, T. A. and Winters, T. E. (1968) *Tetrahedron Letters* 3145.
17. Vokac, K., Samek, Z., Herout, V. and Sorm, F. (1968) *Tetrahedron Letters*, 3855.
18. Vokac, K., Samek, Z., Herout, V. and Sorm, F. (1969) *Coll. Czech. Chem. Commun.* **34**, 2288.
19. Verzar-Petri, G. and Shalaby, A. S. (1977) *Acta Agron. Hung.* **26**, 337.
20. Stahl, E. (1953) *Dtsch. Apoth. Ztg.* **93**, 197.

NEUE EUDESMANOLIDE AUS *GAZANIA KREBSIANA**

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Die südafrikanische Gattung *Gazania*, Tribus Arctotideae, ist chemisch noch wenig untersucht. Bisher sind lediglich Spuren von Acetylenverbindungen [1] und einige Carotinoide [2] isoliert worden. Wir haben jetzt *G. krebsiana* Less. näher untersucht. Die Wurzeln enthalten die Triperpene 1-4 sowie in sehr kleiner Menge zwei Sesquiterpenlactone. Die etwas weniger polare Verbindung mit der Summenformel C₁₅H₁₈O₂ zeigt im ¹H-NMR-Spektrum sehr charakteristische Signale, die nur mit der Konstitution 5 vereinbar sind (s. Tabelle 1), während es sich bei der polareren Substanz um das entsprechende 8α-Isovaleryl-oxy-Derivat 6 handeln muß. Doppelresonanz-Experimente zeigen klar, daß bei 5 das nicht gut aufgelöste Signal bei 5.72 dem 3-H zugeordnet werden muß. Bei Einstrahlung auf das 4-Methylsignal bei 2.00 erhält man ein klares dreifaches

Dublett (*J* = 5.5, 3, 0.7 Hz). Das betreffende Proton koppelt also vicinal mit den 2-H (*J* = 5.5), sowie allyisch mit 5-H (*J* = 3) und 1-H (*J* = 0.7). Derartige Kopplungsmuster sind charakteristisch für entsprechende Cyclohexadiene. Weiterhin kann man zeigen, daß das Signal für 6β-H (*dd* 4.03) mit dem verbreiterten Dublett bei 2.68 und dem *dddd* bei 2.54 koppelt. Letzteres ist nur dem 7α-H zuzuordnen. Entsprechend beobachtet man bei Einstrahlung auf 2.54 eine Entkopplung der Signale für 13-H und eine Veränderung der Multipletts bei 2.03 und 1.62, die zweifellos den Protonen an C-8 zuzuordnen sind. Beim Spektrum von 6 beobachtet man ein *ddd* 5.23 (*J* = 10, 10, 4), sowie eine geringfügige Tieffeldverschiebung der Signale für 5-, 6-, 7- und 13-H. Dieses ist charakteristisch für Lactone mit einem Esterrest in 8α-Stellung [3]. Alle übrigen Signale entsprechen weitgehend denen von 5 (s. Tabelle 1). 5 ist ein 1,2-Dehydro-α-cyclocostunolid, das wir Gazaniolid nennen möchten.

Derartige Verbindungen sind bisher noch nicht als Naturstoff isoliert worden. Evt. Vorstufen von 5 sind jedoch Santamarin [4] bzw. Douglasin [5]. Bemerkens-

* 177. Mitt. in der Serie "Natürlich vorkommende Terpen-Derivate"; 176. Mitt. Bohlmann, F. und Mahanta, P. K. (1979) *Phytochemistry* **18**, 348.